

Gene specific therapy for arrhythmogenic disorders

Silvia G. Priori, Elena Ronchetti, Mirella Memmi

Molecular Cardiology Laboratories, Fondazione Salvatore Maugeri, IRCCS, Pavia, Italy

(Ital Heart J 2000; 1 (Suppl 3): S52-S54)

Address:

Dr.ssa Silvia G. Priori
Cardiologia Molecolare
Fondazione Salvatore
Maugeri, IRCCS
Via Ferrata, 8
27100 Pavia
E-mail: spriori@fsm.it

Introduction

The long QT syndrome (LQTS) is a familial disease characterized by an abnormally prolonged QT interval and by stress-mediated life-threatening ventricular arrhythmias. LQTS usually manifests itself in children and teenagers in the absence of structural cardiac abnormalities¹.

LQTS is not only an important disease for the high mortality rate when misdiagnosed, but it represents a model to understand the complex link between genetically transmitted alterations in cardiac electrophysiology, the autonomic nervous system, and sudden cardiac death. These considerations explain the interest of clinical and experimental research in LQTS, which has progressively grown from the early observations² to the recent development provided by the molecular understanding of the disease.

Clinical presentation and diagnosis

A familial pattern of inheritance is not apparent in approximately 20-25% of cases, which therefore have to be classified as sporadic: these individuals probably have a *de novo* mutation and will become founders of kindred with LQTS³. Alternatively, they may represent the only individuals with the disease phenotype among a family where other non-penetrant gene carriers are present, because of the incomplete penetrance of some mutations⁴. The clinical history is typically represented by episodes of loss of consciousness under emotional or physical stress.

Diagnostic criteria

In quite a few cases the symptoms of LQTS are so characteristic that diagnosis

presents no problems for the physician aware of the disease. Borderline cases do exist and may require evaluation of the several features described above besides clinical history and surface electrocardiogram. A first diagnostic algorithm has been proposed by Schwartz et al.⁵ in 1993. Unfortunately, despite the diagnostic criteria help in unifying requirements for diagnosis, they still leave open the dilemma of borderline cases: the gold standard for diagnosis of LQTS is therefore the molecular identification of the mutation in the DNA.

Toward molecular diagnosis of long QT syndrome

In the last 6 years the extensive study of LQTS families from a molecular biology standpoint has made substantial progress in the understanding of the pathophysiologic mechanisms underlying the LQTS. In 1995 the genes for chromosome 3, 7 and 11 were identified⁶⁻¹⁰ as, *SCN5A*, the cardiac sodium channel, *HERG*, the gene encoding the rapid component of the delayed rectifier (I_{Kr}), and as *KvLQT1* encoding for the membrane ionic channel conducting I_{Ks} , the slow component of the I_K . More recently two additional genes have been identified on chromosome 21: *KCNE1* and *KCNE2*^{11,12}. Overall only 1-2% of patients carry mutations in the latter genes that encode for the auxiliary protein modulating function of *KvLQT1* and *HERG* gene products.

Electrophysiologic characteristics of the mutations identified in long QT syndrome genes. *SCN5A*. The three initially identified^{8,13} mutations provided the rationale for the understanding of LQT3 that still holds

true. These mutations consisted of a 9 base pair deletion (Δ KPQ) and two point mutations (R1644H and N1325S). All these three mutations affect a region important for the inactivation of the Na^+ current. Subsequently, Bennett et al.¹⁴ characterized the Δ KPQ mutation by expressing the altered cardiac sodium channel in *Xenopus oocytes*. Mutant channels showed a sustained inward current during membrane depolarization. Dumaine et al.¹⁵ further characterized the electrophysiologic consequences of the Δ KPQ and the two point mutations showing that all three mutations increased sodium inward current but the defects were of a different severity.

HERG. Sanguinetti et al.¹⁶ expressed the protein coded by *HERG* in *Xenopus oocytes* and demonstrated that *HERG* encodes the rapid component of the delayed rectifier. The electrophysiologic properties of expressed *HERG* were nearly identical to the I_{Kr} current in cardiac myocytes. The same authors¹⁷ demonstrated that *HERG* mutations produce a loss of channel function resulting in a reduced repolarizing K^+ current that may account for the clinical phenotype.

KvLQT1. Wang et al.¹⁰ have identified the gene responsible for chromosome 11-linked LQTS (LQT1), specifically *KvLQT1*. Ten different missense mutations and intragenic deletions in *KvLQT1* were found by the authors in 16 LQTS families. *KvLQT1* proteins co-assemble with the *KCNE1* gene product to form the functional I_{Ks} conducting channel¹¹. Several expression studies of *KvLQT1*-minK mutations have been performed and they showed a reduction of the I_{Ks} current. Since I_{Ks} is the most important current that allows shortening of repolarization at faster rates (i.e. during adrenergic activation) it seems logical to speculate that LQT1 individuals are more vulnerable during conditions of high adrenergic activity. Recent data by Schwartz (personal communication) have confirmed that most of the arrhythmic events in LQT1 patients occur during physical and emotional stress.

Therapy

Independently of the primary abnormality involved in its pathogenesis, the trigger for arrhythmias in most LQTS patients is a sudden sympathetic discharge¹. Antiadrenergic interventions therefore represent the most logical therapeutic approach. Other therapeutic approaches (i.e. pacemakers, implantable cardioverter defibrillators, and non-antiadrenergic pharmacological therapies) have been also proposed. The most appealing challenge for the near future however lies in the attempt to design pharmacological interventions that may counteract the consequences of the individual mutations. Data are still preliminary, however a few strategies have been highlighted as discussed below.

Experimental basis for gene specific therapy. Based on the evidence that LQT3 is caused by alterations in the inactivation of cardiac sodium channels^{14,15} and that LQT2 is caused by a reduction in the delayed rectifier potassium current¹⁷, we attempted to develop the first cellular model for LQTS¹⁸. Our goal was to provide a means for assessing the effect of different interventions in two forms of the disease: LQT2 and LQT3.

We exposed guinea pig ventricular myocytes¹⁸ to anthopleurin, a toxin that interferes with the inactivation of I_{Na} , and to dofetilide, a selective blocker of I_{Kr} in order to obtain a prolongation of cellular repolarization. Both anthopleurin and dofetilide induced a prolongation of action potential duration confirming that either a reduced I_{Kr} or a persistent inward sodium current can cause the classic phenotypic alteration of LQTS, i.e. prolongation of the QT interval. Using this experimental preparation mimicking LQT3 and LQT2, respectively, we tried to differentiate the phenotypic manifestations of these two forms of the disease by characterizing the response to the sodium channel blocker mexiletine and to rapid pacing.

Exposure to the sodium channel blocker mexiletine significantly reduced the action potential duration in cells treated with anthopleurin while it did not modify the prolongation induced by dofetilide. This demonstrated that it was possible to normalize the action potential duration by blocking the persistent inward sodium current in the LQT2 mimicking model. This finding fits in very well with the demonstration by Dumaine et al.¹⁵ that mexiletine could reverse the changes induced by the three LQT3 mutations on *SCN5A*.

When fast pacing was performed anthopleurin treated cells showed a prompt shortening of action potential duration toward control values. Overall, these data demonstrated that pharmacological interventions aimed at mimicking phenotypic manifestations of LQT3 and LQT2 at a cellular level, result in differential responses to sodium channel blockade, and to rapid pacing. From a clinical standpoint these results might indicate that LQT3 patients could favorably respond to sodium channel blockade and to pacing that would prevent bradycardia-related QT prolongation.

Gene specific therapy for long QT syndrome patients. Based on this experimental evidence, we tested the hypothesis that mexiletine and physiologically-induced increases in heart rate would shorten the QT interval more in LQT3 than in LQT2 patients¹⁹.

Fifteen LQTS patients who were referred to our center entered the study and were characterized genetically. Eight patients belonged to three LQTS families genetically linked to chromosome 3 (LQT3), while 7 patients belonged to two families linked to chromosome 7 (LQT2). Mexiletine significantly shortened the QT interval among LQT3 patients but not among LQT2 patients. When we examined the response to increases in heart rate, we found that LQT3

Table I. Proposed algorithm for gene specific therapy in long QT syndrome.

Genetic variant	Gene specific treatment	Rationale
LQT1	Antiadrenergic	The defective current I_{Ks} is important during adrenergic activation, antiadrenergic interventions reduce the effects of its dysfunction
LQT2	Potassium supplement Potassium sparing diuretics	The I_{Kr} current defective in these patients is augmented by increased extracellular potassium
LQT3	Sodium channel blockers	The genetic defect causes an excess of inward sodium current

patients shortened their QT interval in response to heart rate changes more than LQT2 patients and more than the healthy controls.

Conclusions

Even if data on gene specific therapy in LQTS should still be considered at a preliminary phase of development, they support important clinical considerations. LQT3 patients may not be at a particularly high risk during physical stress because during progressive sinus tachycardia, their QT intervals would markedly shorten. They might benefit less than other LQTS patients from antiadrenergic therapy while they might benefit from chronic therapy with mexiletine and pacing. On the other hand, LQT1 patients are more likely to be at risk for cardiac arrest under stressful conditions, because the arrhythmogenic effect of catecholamines would be enhanced by the lack of appropriate QT shortening when heart rate increases. LQT1 patients are likely to be protected by antiadrenergic therapy. In LQT2 patients an increase in the extracellular concentration of potassium may shorten the QT interval²⁰; it is therefore possible that this subgroup of individuals may benefit from combined therapy with potassium sparing diuretics, oral potassium supplements and beta-blockers. Overall the proposed algorithm for gene specific therapy in LQTS is outlined in table I.

References

- Schwartz PJ, Priori SG, Napolitano C. Long QT syndrome. In: Zipes DP, Jalife J, eds. Cardiac electrophysiology. From cell to bedside. 3rd edition. Philadelphia, PA: WB Saunders, 2000: 597-615.
- Schwartz PJ, Periti M, Malliani A. The long Q-T syndrome. Am Heart J 1975; 89: 378-90.
- Schwartz PJ. Idiopathic long QT syndrome: progress and questions. Am Heart J 1985; 109: 399-411.
- Priori SG, Napolitano C, Schwartz PJ. Low penetrance in the long QT syndrome. Clinical impact. Circulation 1999; 99: 529-33.
- Schwartz PJ, Moss AJ, Vincent GM, Crampton RS. Diagnostic criteria for the long QT syndrome: an update. Circulation 1993; 88: 782-4.
- Keating MT, Atkinson D, Dunn C, Timothy K, Vincent GM, Leppert M. Linkage of a cardiac arrhythmia, the long QT syndrome, and the Harvey ras-1 gene. Science 1991; 252: 704-6.
- Jiang C, Atkinson D, Towbin JA, et al. Two long QT syndrome loci map to chromosomes 3 and 7 with evidence for further heterogeneity. Nat Genet 1994; 8: 141-7.
- Wang Q, Shen J, Splawski I, et al. SCN5A mutations associated with an inherited cardiac arrhythmia, long QT syndrome. Cell 1995; 80: 805-11.
- Curran ME, Splawski I, Timothy KW, Vincent GM, Green ED, Keating MT. A molecular basis for cardiac arrhythmia: HERG mutations cause long QT syndrome. Cell 1995; 80: 795-803.
- Wang Q, Curran ME, Splawski I, et al. Positional cloning of a novel potassium channel gene: KvLQT1 mutations cause cardiac arrhythmias. Nat Genet 1996; 12: 17-23.
- Splawski I, Tristani-Firouzi M, Lehmann MH, Sanguinetti MC, Keating MT. Mutations in the hminK gene cause long-QT syndrome and suppress I_{Ks} function. Nat Genet 1997; 17: 338-40.
- Abbott GW, Sesti F, Splawski I, et al. MiRP1 forms I_{Kr} potassium channels with HERG and is associated with cardiac arrhythmia. Cell 1999; 97: 175-87.
- Wang Q, Shen J, Li Z, et al. Cardiac sodium channel mutations in patients with long QT syndrome, an inherited cardiac arrhythmia. Hum Mol Genet 1995; 4: 1603-7.
- Bennett PB, Yazawa K, Makita N, George AL Jr. Molecular mechanism for an inherited cardiac arrhythmia. Nature 1995; 376: 683-5.
- Dumaine R, Wang Q, Keating MT, et al. Multiple mechanisms of Na⁺ channel-linked long-QT syndrome. Circ Res 1996; 78: 916-24.
- Sanguinetti MC, Jiang C, Curran ME, Keating MT. A mechanistic link between an inherited and an acquired cardiac arrhythmia: HERG encodes the I_{Kr} potassium channel. Cell 1995; 81: 1-20.
- Sanguinetti MC, Curran ME, Spector PS, Keating MT. Spectrum of HERG K⁺-channel dysfunction in an inherited cardiac arrhythmia. Proc Natl Acad Sci USA 1996; 93: 2208-12.
- Priori SG, Napolitano C, Cantò F, Brown AM, Schwartz PJ. Differential response to Na⁺ channel blockade, β -adrenergic stimulation, and rapid pacing in a cellular model mimicking the SCN5A and HERG defects present in the long QT syndrome. Circ Res 1996; 78: 1009-15.
- Schwartz PJ, Priori SG, Locati EH, et al. Long QT syndrome patients with mutations on the SCN5A and HERG genes have differential responses to Na⁺ channel blockade and to increases in heart rate. Implications for gene-specific therapy. Circulation 1995; 92: 3381-6.
- Compton SJ, Lux RL, Ramsey MR, et al. Genetically defined therapy of inherited long-QT syndrome. Correction of abnormal repolarization by potassium. Circulation 1996; 94: 1018-22.